



Fluorescence aptasensor based on competitive-binding for human neutrophil elastase detection

Jing-Lin He, Zai-Sheng Wu, Song-Bai Zhang, Guo-Li Shen, Ru-Qin Yu*

State Key Laboratory for Chemo/Biosensing and Chemometrics, College of Chemistry and Chemical Engineering, Hunan University, Changsha 410082, PR China

ARTICLE INFO

Article history:

Received 6 June 2009

Received in revised form 8 September 2009

Accepted 13 September 2009

Available online 19 September 2009

Keywords:

Aptasensor

Competitive-binding reaction

Human neutrophil elastase

Molecular beacon

Fluorescence

ABSTRACT

To our knowledge, we report the first fluorescence aptasensor for detecting human neutrophil elastase (HNE) in homogeneous solution. The biosensor contains a short DNA scrambled sequence strand (SS) complementary to part of the aptamer sequence or the loop of molecular beacon (MB). The aptamer-HNE recognition event involves competition between the molecular beacon and loose HNE aptamer for the binding the short DNA strand. The new biosensor can detect as little as 0.34 nM of HNE, and the response is linear in the tested concentration range of 0.34–68 nM with the detection limit of 47 pM.

© 2009 Elsevier B.V. All rights reserved.

1. Introduction

Neutrophil granulocytes are primary antimicrobial effector cells of the innate immune system and contribute to the first line of defence against infectious agents or nonself substances that penetrate the body's physical barriers [1]. The human neutrophil elastase (HNE), a serine proteinase, is the principal enzyme released from neutrophils [2]. It is capable of solubilising fibrous elastin, cartilage proteoglycans, several collagens and fibronectin, hence facilitating the migration of cells to inflammatory regions. HNE is thought to physiologically participate in disease resistance by facilitating the degradation and phagocytosis of pathogenic bacteria [3]. Uncontrolled activity of HNE has been shown to contribute to the pathogenesis of rheumatoid arthritis [4], chronic obstructive pulmonary disease [5], adult respiratory distress syndrome [6], glomerulonephritis [7], and chronic and burn wounds [8,9].

Lin et al. [10] identified a specific high-affinity DNA ligand targeted to HNE using an *in vitro* selection technique (SELEX). The DNA ligand folds into a G-quartet structure with duplex ends. According to Davis's work [11], the fluoresceinated DNA ligand equally effective as an anti-HNE antibody in detecting HNE can be useful in diagnostic applications based on flow cytometry. Charlton et al. [12] applied the DNA aptamer ligand to the field of diagnostic imaging.

Fluorescence spectroscopic methods offer high sensitivity and selectivity and applicability in nonseparation detection and *in situ* monitoring [13–16]. Nutiu and Li [17], for example, described aptamer-based fluorescent reporters for detecting for thrombin over the range of 10 nM–1.0 μ M that a structural switch releases a dabcyt-labeled nucleotide strand from the fluorophore-labeled aptamer. Li et al. [18], on the other hand, prepared a molecular aptamer beacon (MAB) for recognizing the thrombin producing significant fluorescence signal change, which is attributed to a significant conformational change at low concentration (112 pM). Lerga and O'Sullivan [19] developed thrombin MAB labeling fluorescein and coumarin to selectively detect Ca^{2+} and Mg^{2+} with detection limit (40 μ M) for determination of total water hardness in tap and bottle water. Heyduk and Heyduk [20] introduced a fluorescent assay involving thrombin-induced coassociation of two aptamers with analyte resulted in bringing the two short fluorophore-labeled oligonucleotides into proximity. Change of fluorescence resonance energy transfer was even at the lowest thrombin concentration tested (50 pM).

In our research, we proposed a sensitive method for HNE analysis in homogeneous solution. The molecular beacon was designed by appending fluorescein to 5'-ends and dabcyt quencher to 3'-ends. The HNE aptamer was free-labeled in order to retain the bioactivity. The short DNA scrambled sequence was complementary to the binding sites of the HNE aptamer or the loop of molecular beacon. Our biosensor involves aptamer-HNE recognition, competition between the molecular beacon and loose HNE aptamer for binding the short DNA strand, and monitoring the extent of

* Corresponding author. Fax: +86 731 8822782.

E-mail address: rquy@hnu.cn (R.-Q. Yu).

competition through highly sensitive fluorescence detection of the captured HNE.

2. Experimental

2.1. Materials and reagents

HNE from human plasma (SERVA Electrophoresis GmbH, Heidelberg, Germany) was dissolved in a storage buffer (50 mM Tris-HCl, 50 mM NaCl, 3.0 mM KCl, 1.0 mM MgCl₂, pH 7.5) to yield 3.39 μM solutions. This standard was divided into 100 μL aliquots and stored at -20 °C. Immediately before use, each aliquot was warmed to 4 °C and diluted with a storage buffer solution to the required concentration. HNE concentration mentioned in our work was the molarity of HNE storage solution before fluorescence measurements.

Oligonucleotides used in the study were customer-designed and synthesized by Sangon Biotechnology Co. Ltd. (Shanghai, China) as follows.

The HNE aptamer: 5'-TAGC GATA CTGC GTGG GTTG GGGC GGGT AGGG CCAG CAGT CTCGT,

The molecular beacon (MB): 5'-fluorescein-CCTA GCGC GTGG GTTG GGGC TAGG-Dabcyl,

Four scrambled sequence (SS):

S12: 5'-CCCA ACCC ACGC; S14: 5'-GC CCCA ACCC ACGC; S16: 5'-CCGC CCCA ACCC ACGC; S19: 5'-TAC CCGC CCCA ACCC ACGC.

Ultrapure water used to prepare all of the solutions was obtained through a Nanopure Infinity Ultrapure water system (Barnstead/ThermoFisher Corp., Dubuque, IA) with an electrical resistance larger than 18.3 MΩ. All buffer solutions and ultrapure water were sterilized and used throughout experiments. Sodium chloride, potassium chloride, human serum albumin (HSA), magnesium chloride and Tris-HCl were purchased from China National Medicines Co. Ltd. (Beijing, China).

2.2. Fluorescence measurements

If not otherwise specified, the following concentrations of oligonucleotides were used for fluorescence measurements: 40 nM for aptamers, 40 nM for the SS and 40 nM for the MB. MB was dissolved in storage buffer to form hairpin duplex, while HNE aptamer and SS were dissolved in ultrapure water to keep flexible structure. The binding buffer (pH 7.5) consisted of 100 mM Tris-HCl, 850 mM NaCl, 16 mM KCl, 5.0 mM MgCl₂ and 0.12% HSA (wt.%).

To perform the fluorescence experiments, 50 μL HNE solution of a specific concentration and 100 μL binding buffer were incubated

with 50 μL HNE aptamer for 45 min. After the addition of 50 μL of a SS solution, the mixture was incubated for 10 min at 10 °C and then added 50 μL MB solution.

All fluorescence measurements were performed in 50 mM Tris-HCl (pH 7.5), 300 mM NaCl, 6.0 mM KCl, 2.0 mM MgCl₂ and 0.02% HSA (wt.%). All fluorescence spectra were recorded on a Hitachi F-4500 fluorescence spectrometer (Hitachi Ltd., Japan) controlled by FL Solution software to fit a curve and determine the peak height. A quartz fluorescence cell with an optical path length of 1.0 cm was used. The excitation was made at 495 nm with a recording emission range of 500–600 nm. All excitation and emission slits were set at 5 nm. The fluorescence spectrum of all mixtures was recorded at 10 ± 2 °C.

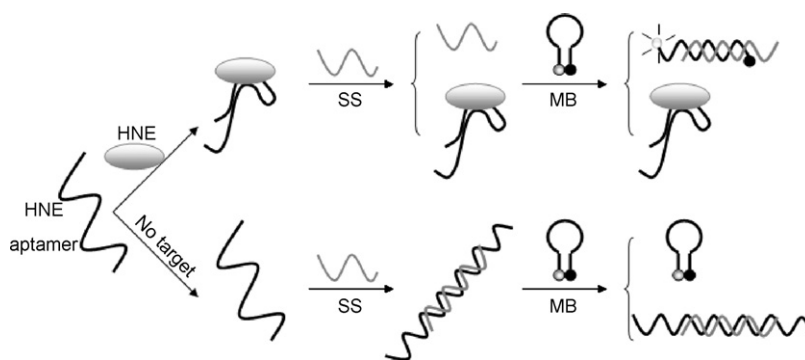
3. Results and discussion

3.1. Experimental principle

Scheme 1 illustrates the concept of the competitive-binding process coupled with fluorescence increase in this design. Before the addition of target, HNE aptamer (the long black coil) kept flexible structure in the blank experiment. SS (the short grey coil) was introduced in this homogeneous solution, and naturally binds to the aptamer. When MB (the short coil with two rotundities) was introduced, the hybridization of SS with MB would not take place immediately, which kept the fluorescence off. In the presence of HNE (the ellipse), the flexible structure of aptamer was transformed into a HNE-aptamer duplex assembly with G-quartet structure. Upon addition of SS and MB, the corresponding complementary sections of SS would hybridize with MB producing a strong concentration-dependent fluorescent signal.

3.2. Fluorescence increase of HNE binding

Fig. 1 demonstrates fluorescence spectra of MB solutions, a mixture of aptamer/SS/MB, an assembly of aptamer/HNE/SS/MB and a duplex of SS/MB. The sample containing MB only (Fig. 1, curve a) shows rather low fluorescence intensity for its hairpin structure keeping the fluorophore and the quencher into proximity. A slight fluorescence increase was observed when the MB was added into solution containing aptamer and SS (curve b). It can be interpreted as that a small quantity of MB captured SS dissociated from aptamer/SS. In the presence of HNE (17 nM, curve c), a significant fluorescence intensity increase (102%) was recorded at the emission peak around 518 nm. Binding of target proteins forces the aptamers to undergo a conformational change, while MB opens its hairpin structure leading to an increase in fluorescent signal. When



Scheme 1. Schematic representation of the aptamer-based HNE sensor. HNE: the ellipse; unmodified aptamer: the long coil; DNA scrambled sequence (SS): the short coil; DNA molecular beacon (MB): the short coil with two rotundities.

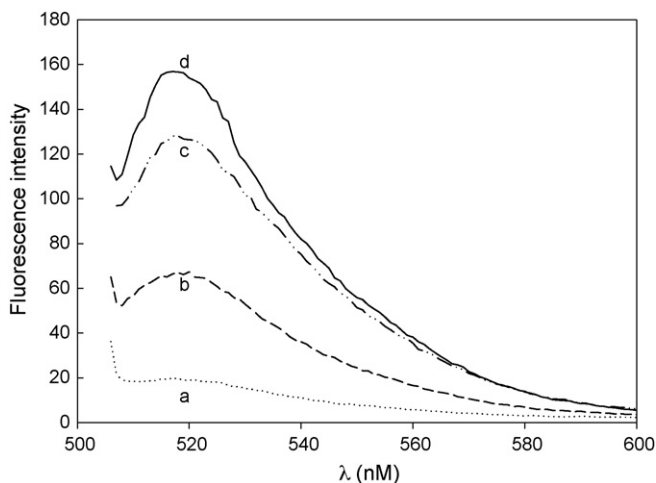


Fig. 1. Fluorescence spectra of solutions containing MB (a), a mixture of aptamer, SS and MB (b), a mixture of aptamer/HNE, SS and MB (c), and a complex of SS/MB (d). The relative standard deviations obtained by five repeated measurements were not more than 5%.

binding buffer contains SS and MB without addition of aptamer (curve d), the fluorescence intensity recorded was the maximum fluorescence intensity obtainable corresponding to the complete hybridization of MB with SS. This observation shows that SS could hybridize really with MB inducing fluorescence increase and HNE could be detected using the proposed method.

3.3. Experimental conditions of target detection

The efficiency of the competitive-binding process depends on the amount of SS released from aptamer and the ability of SS to force the MB stem apart. To investigate the effect of SS strand length on the performance of this method, the aptamer and aptamer/HNE assembly were hybridized with specific SS with a 12-, 14-, 16- or 19-nucleotide segment. As shown in Fig. 2, S12 provides the lowest fluorescent signal in the presence of HNE (curve a') and also the least background (curve a). S12 has the least number of complementary oligonucleotides, which are too short to promote efficient hybridization with MB. The number of adjacent nucleotides on aptamer-SS binding region was 14 in the assay with S14, S16 or S19. The extra -CG sequence was designed to hybridize with the stem region of MB. The extra sequence might be beneficial to increase the ability of the corresponding SS to force the MB stem apart and restore the fluorescence. When the number of nucleotides rose from 14, 16 to 19, the decreasing background fluorescence in absence of HNE (curves b, c and d) seemed due to the ever increasing hybridizing force between the aptamer and the corresponding SS strand. In presence of HNE (curve b', c' and d'), a longer SS strand would induce higher fluorescence increase. In particular, S19 provides the best signal increase for HNE sensing. The extra length of the S19 strand (5'-TAC sequence) might loose the inner molecular interaction between the MB stem sections, which probably lead to fluorescence signal increase. Therefore, S19 was used as optimum SS in the experiment.

We also investigated the effects of pH on the response of the proposed sensor to HNE over the pH range from 7.0 to 9.0 and

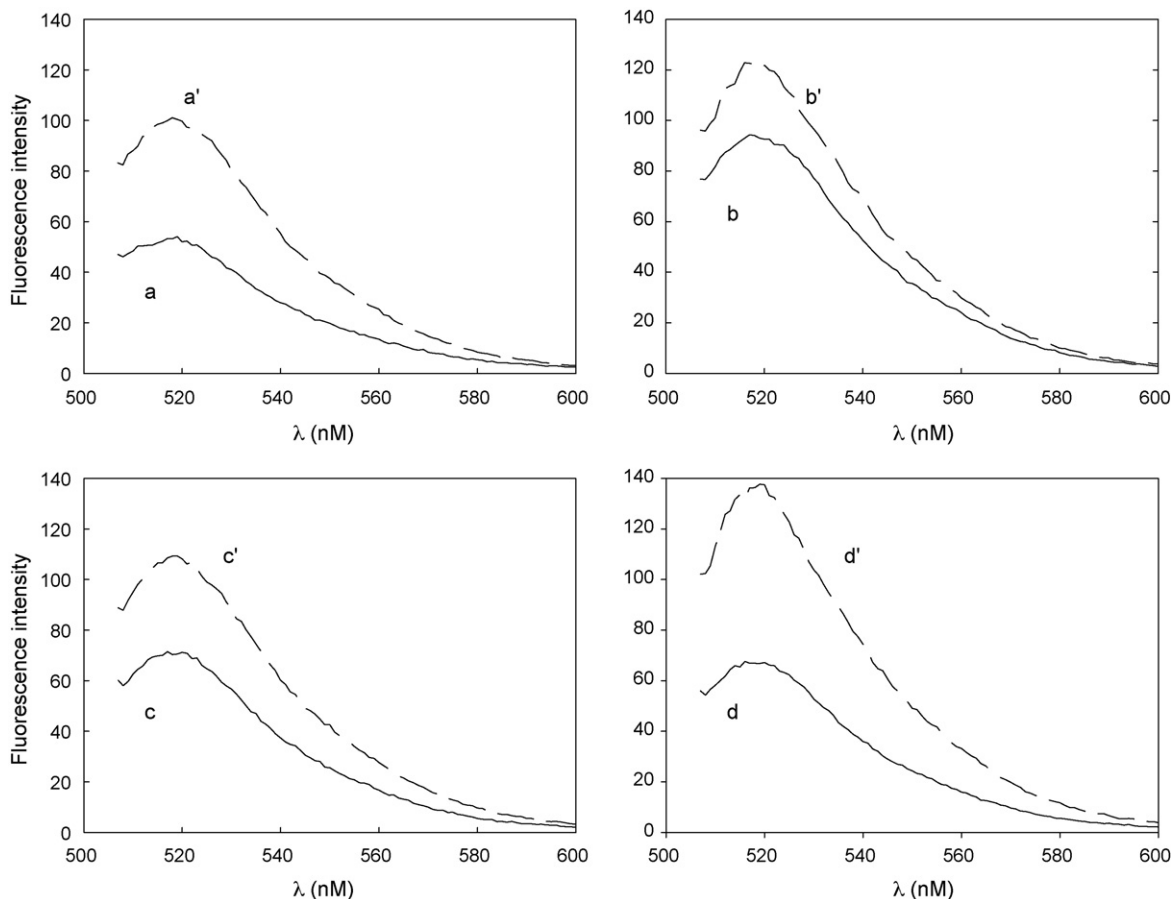


Fig. 2. Fluorescence spectra of the sensor using S12 (a and a'), S14 (b and b'), S16 (c and c') and S19 (d and d') with and without HNE respectively. The relative standard deviations obtained by five repeated measurements were not more than 5%. S12: 5'-CCCA ACCC ACCG; S14: 5'-GCCC CAAC CCAC GC; S16: 5'-CCGC CCCA ACCC ACCG; S19: 5'-TACC CGCC CCAA CCGC.

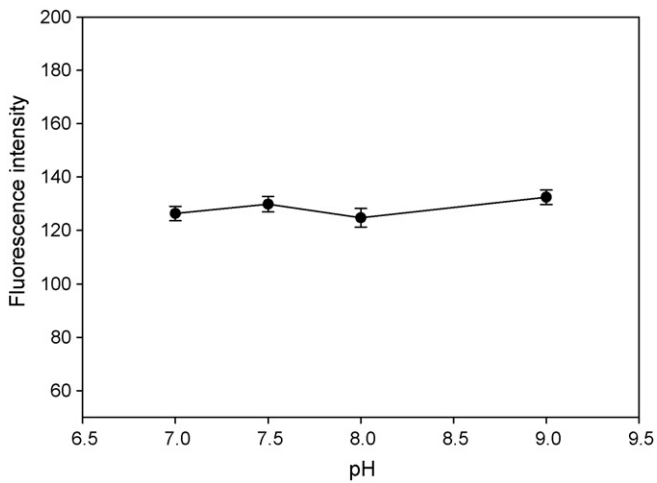


Fig. 3. The effect of pH of the incubating solution on the activity of the sensor containing HNE (17 nM). The standard deviations obtained by five repeated measurements were shown as the error bars.

the results are shown in Fig. 3. As can be seen, the sensor can be carried out in this pH range. There is no obvious change in the fluorescence of the assay when the pH of the buffer increases from 7.0 to 9.0. This indicates that competitive-binding reaction can occur over a wide pH range. An ideal pH of 7.5 was chosen for the sensor experiments, which is also a general pH value in immunology.

Thermal denaturation experiments were performed (Fig. 4) to investigate the relationship between fluorescence signal and incubation temperature. For this purpose, we increased the incubation temperature from 10 to 24, 37 or 45 °C. The binding event of aptamer/S19 is less efficient at high temperatures in the absence of HNE (Fig. 4, closed circles), resulting in the increase in fluorescence background. The low fluorescence intensity at 45 °C in absence or presence of HNE was possible due to the thermal quenching of the fluorescence. After introducing the analyte (Fig. 4, open circles), a high incubation temperature might lead to the HNE protein denaturation. As a result, more aptamer dissociated from aptamer/HNE conjugate and hybridized with S19, bringing fluorescence intensity to decrease. This phenomenon indicated that the optimum incubation temperature was 10 °C.

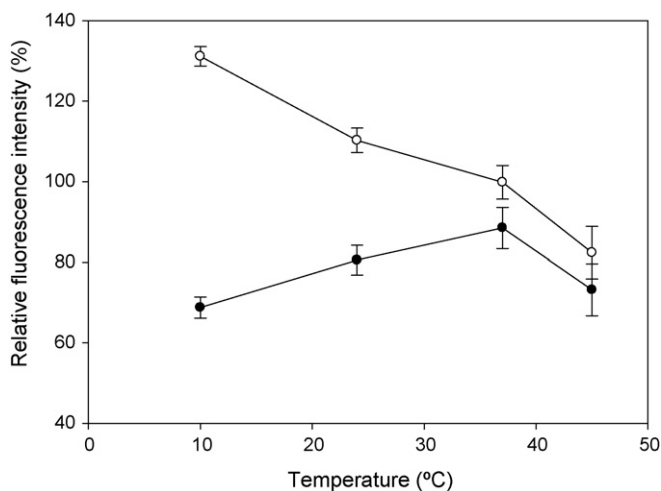


Fig. 4. The DNA aptamer mixture was incubated at 10, 24, 37 and 45 °C with (open circles) and without (closed circles) 17 nM HNE. The standard deviations obtained by five repeated measurements were shown as the error bars.

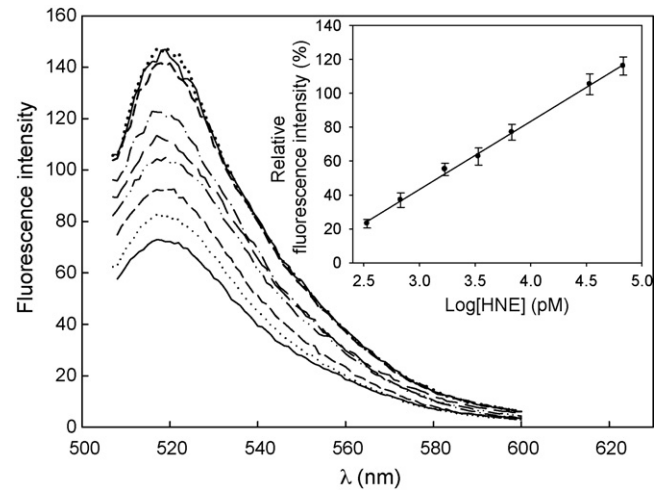


Fig. 5. Fluorescence spectra of sensors in the presence of HNE at different concentrations (from bottom to top): (1) 0.067 nM; (2) 0.34 nM; (3) 0.68 nM; (4) 1.7 nM; (5) 3.4 nM; (6) 6.8 nM; (7) 34 nM; (8) 68 nM; (9) 110 nM. Inset shows the linear fluorescence responses to the logarithm of HNE concentration in the range of 0.34–68 nM. The illustrated error bars represent the standard deviation of three measurements obtained at each HNE concentration.

3.4. Analytical performance of the aptamer-based sensor

Fig. 5 shows the change in fluorescence signal as a function of the HNE concentration. With the increased amount of HNE, the fluorescence intensity of the aptamer probe is increased. The relative fluorescence intensity R (%) was calculated from the equation $R(\%) = ((F_{\text{target}} - F_{\text{no target}}) / F_{\text{no target}}) \times 100\%$, where F is the fluorescence intensity, and the subscripts “target” and “no target” refer to the case in the presence and absence of HNE respectively. Significantly, the aptamer-based sensor exhibited a high degree of sensitivity for HNE (Fig. 5, inset). And the response is linear in the tested concentration range of 0.34–68 nM ($r = 0.9980$). The linear regression equation was expressed as

$$\text{Relative fluorescence intensity (\%)} = 40.12 \log[\text{HNE}] - 77.09$$

The detection limit (at $S/N = 3$) calculated from the slope of the calibration curve in the lowest concentration range was 47 pM.

3.5. Specificity of the HNE fluorescence sensor

Due to the inherent specificity of the aptamer toward its target protein, the fluorescence change of the HNE biosensor is highly

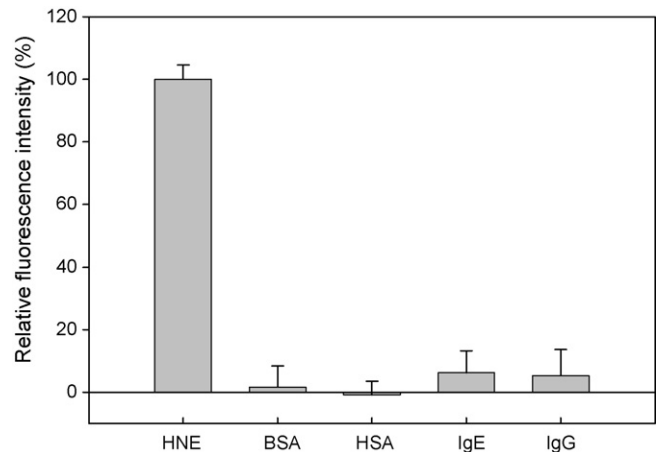


Fig. 6. Selectivity for the proposed sensor of five proteins. The sensor was exposed to HNE (17 nM) or other four proteins (BSA, HSA, IgE and IgG, 85 nM) respectively. Error bars were estimated from at least three independent measurements.

selective. Aliquots of the aptamer was incubated with either HNE (17 nM) or some other proteins commonly present in serum such as bovine serum albumin (BSA), human serum albumin, immunoglobulin E (IgE) and immunoglobulin G (IgG). The relative responses to fluorescence changes of the interfering protein, even at 5-fold higher concentrations (85 nM) than those used for HNE, are negligible in comparison to those from HNE (seen in Fig. 6). Hence the developed biosensor could exhibit a high degree of selectivity for the HNE detection, as expected.

4. Conclusion

We have developed a fluorescence aptasensor for the detection of protein HNE in homogeneous solution using MB. The proposed sensor was designed to observe its fluorescence intensity increase, which was attributed to the difference in structure-switching between the free aptamer and its HNE binding complex. The proposed strategy has been proved applicable in HNE aptasensing. This is of paramount importance for a new selective and highly sensitive method of HNE detection.

Acknowledgements

This research was supported by the National Natural Science Foundation of China (Grants No. 20675028 and 20775023),

the “973” National Basic Research Program of China (No. 2007CB310500) and the Science Commission of Hunan Province.

References

- [1] C. Nathan, *Nat. Rev. Immunol.* 6 (2006) 173.
- [2] N.D. Burg, M.H. Pillinger, *Clin. Immunol.* 99 (2001) 7.
- [3] P. Birrer, *Agents Actions Suppl.* 40 (1993) 3.
- [4] L. Ekerot, K. Ohlsson, *Adv. Exp. Med. Biol.* 167 (1984) 335.
- [5] S.D. Shapiro, *Biochem. Soc. Trans.* 30 (2002) 98.
- [6] C. Jaffray, J. Yang, G. Carter, C. Mendez, J. Norman, *Surgery* 128 (2000) 225.
- [7] T. Oda, O. Hotta, Y. Taguma, H. Kitamura, K. Sudo, I. Horigome, S. Chiba, N. Yoshizawa, H. Nagura, *Hum. Pathol.* 28 (1997) 720.
- [8] D.R. Yager, S.M. Chen, S.I. Ward, O.O. Olutoye, R.F. Diegelmann, I.K. Cohen, *Wound Repair Regen.* 5 (1997) 23.
- [9] F. Grinell, M. Zhu, *J. Invest. Dermatol.* 103 (1994) 155.
- [10] Y. Lin, A. Padmapriya, K.M. Morden, S.D. Jayasena, *Proc. Natl. Acad. Sci.* 92 (1995) 11044.
- [11] K.A. Davis, B. Abrams, Y. Lin, S.D. Jayasena, *Nucleic Acids Res.* 24 (1996) 702.
- [12] J. Charlton, J. Sennello, D. Smith, *Chem. Biol.* 4 (1997) 809.
- [13] M.U. Kumke, G. Li, C.P. Linn, G.T. Walker, L.B. McGown, *Anal. Chem.* 67 (1995) 3945.
- [14] X. Fang, J.J. Li, W. Tan, *Anal. Chem.* 72 (2000) 3280.
- [15] C.J. Yang, H. Lin, W. Tan, *J. Am. Chem. Soc.* 127 (2005) 12772.
- [16] L. Wang, C.J. Yang, C.D. Medley, S.A. Benner, W. Tan, *J. Am. Chem. Soc.* 127 (2005) 15664.
- [17] R. Nutiu, Y. Li, *J. Am. Chem. Soc.* 125 (2003) 4771.
- [18] J.J. Li, X. Fang, W. Tan, *Biochem. Biophys. Res. Commun.* 292 (2002) 31.
- [19] T.M. Lerga, C.K. O'Sullivan, *Anal. Chim. Acta* 610 (2008) 105.
- [20] E. Heyduk, T. Heyduk, *Anal. Chem.* 77 (2005) 1147.